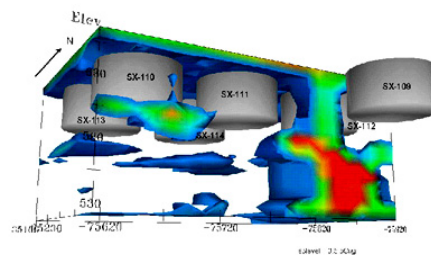
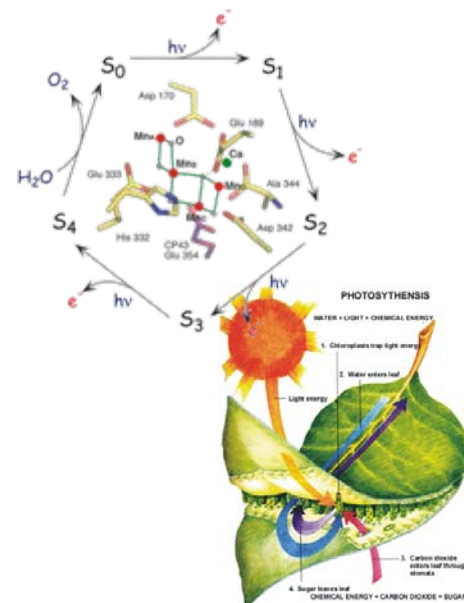
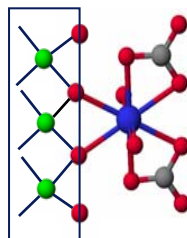
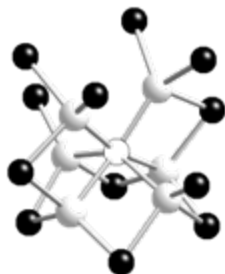


# *X-Ray Absorption Spectroscopy: Practical Aspects*

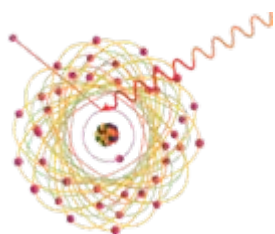
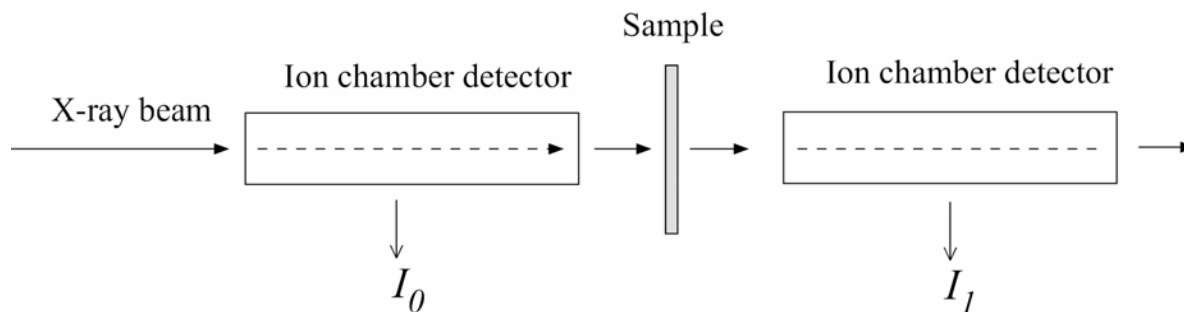


**John Bargar, May 20, 2008**

*SSRL School on Synchrotron X-ray Absorption Spectroscopy  
Techniques in Materials and Environmental Sciences: Theory and  
Application*

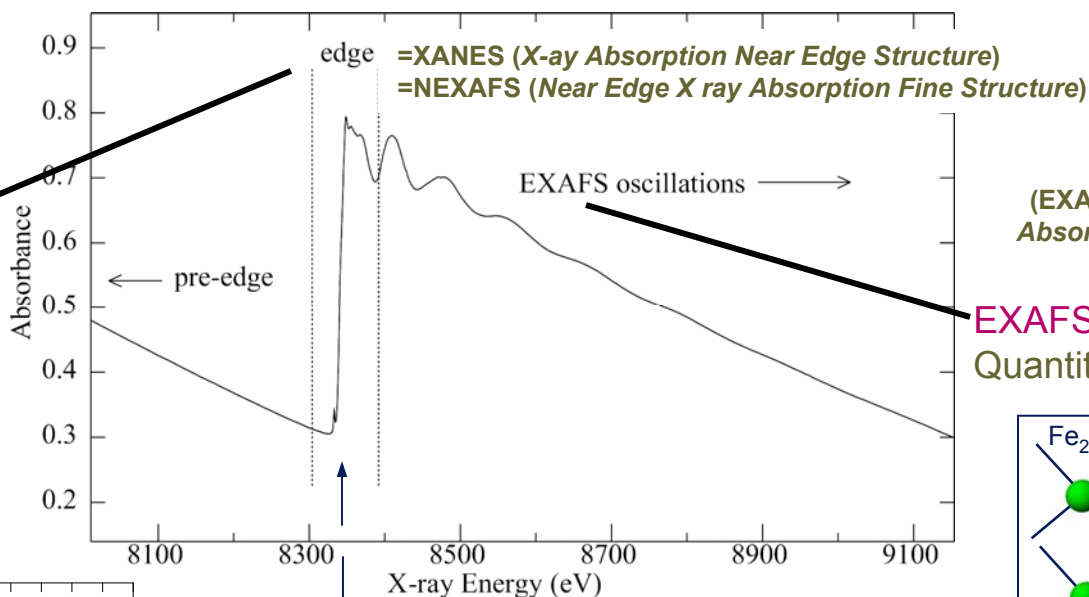
# XAS: What you get out of the measurement:

## Basic Experiment :



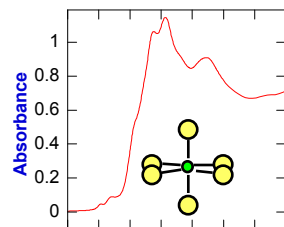
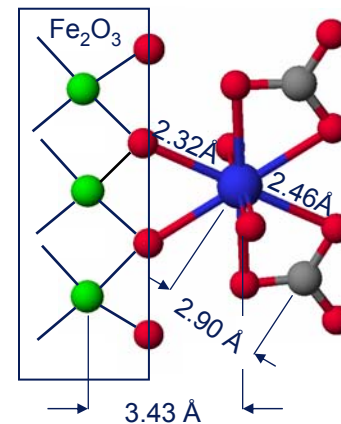
## XANES / NEXAFS

Oxidation state,  
Molecular structure,  
Electronic structure.

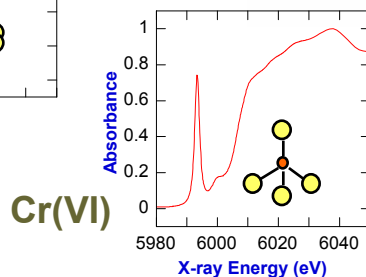


(EXAFS = *Extended X ray Absorption Fine Structure*)

**EXAFS**  
Quantitative Local Structure.

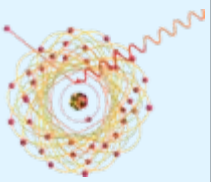


**Cr(III)**



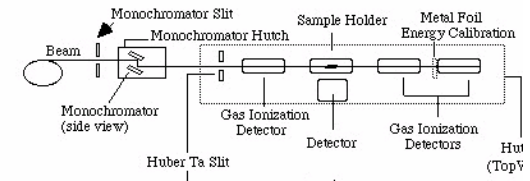
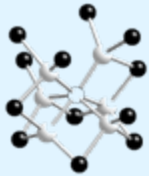
**Cr(VI)**

# Synchrotron-Based Techniques: Key Advantages

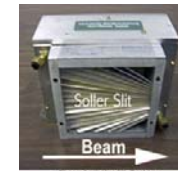
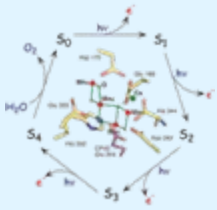


## Three Major Categories

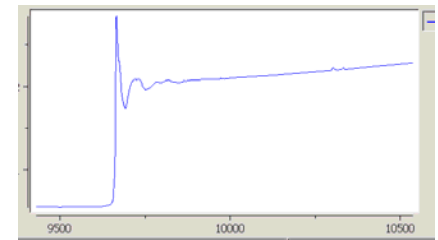
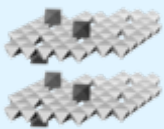
### I. Set-up and optimization of beam lines

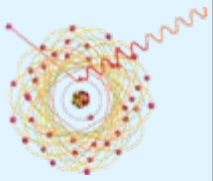


### II. Sample optimization & choice of detector



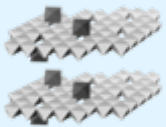
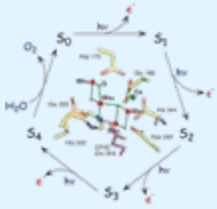
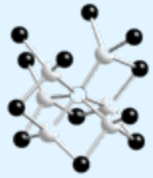
### III. Data Acquisition



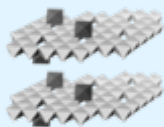
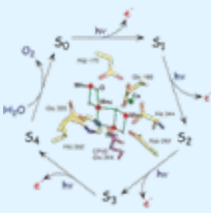
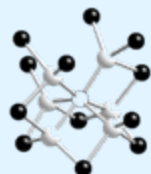
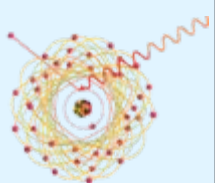
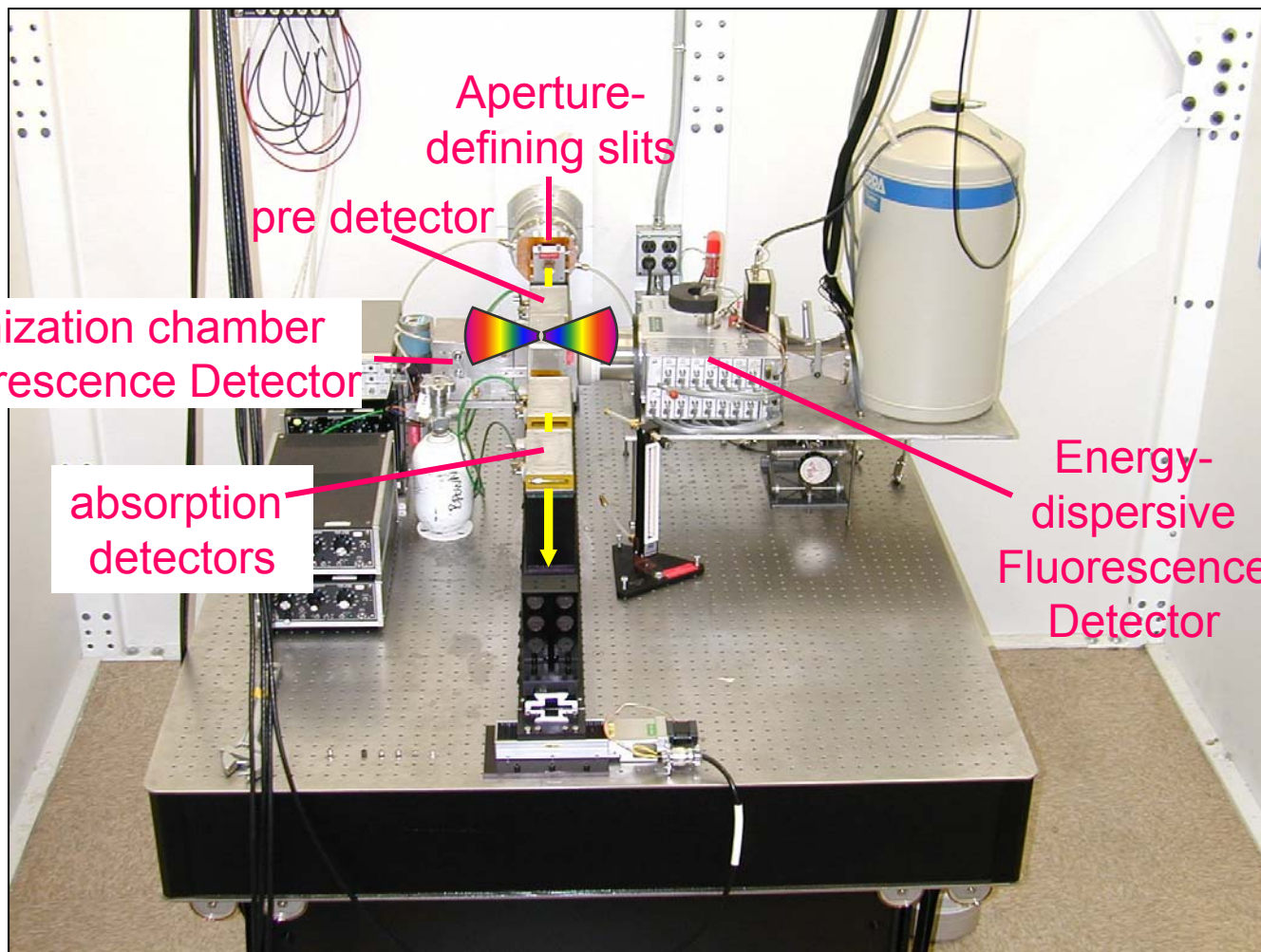


# I. Beam line set-up and optimization

- Major elements of in-hutch equipment
- Major elements outside of hutch
- Ion chambers and their output signal chain
- Mono tuning - why, how, and how much?
- Slit size for samples, impact on resolution
- Energy calibration: why, how, how frequently?

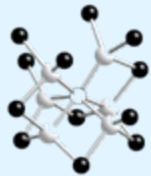
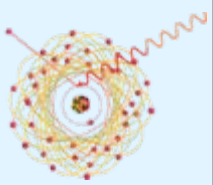


# I. Beam line set-up and optimization: In-hutch instrumentation

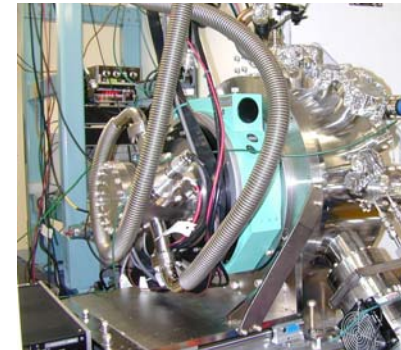




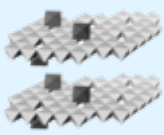
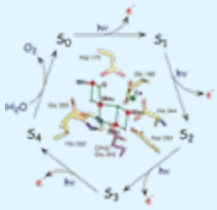
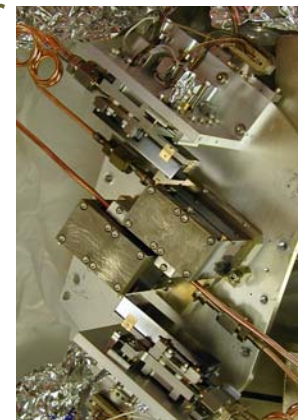
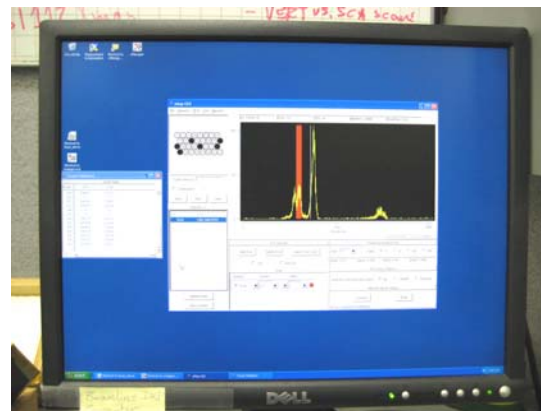
# I. Beam line set-up and optimization: Out-of-hutch instrumentation



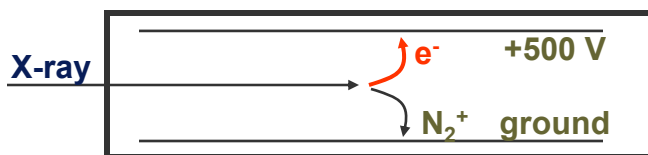
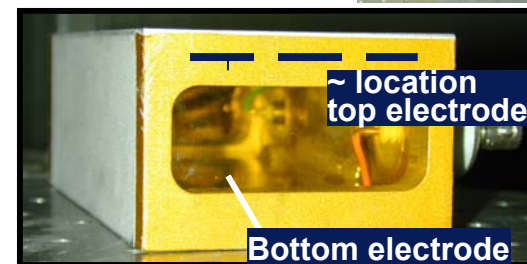
SSRL BL 11-2



Monochromator



# I. Beam line set-up and optimization: Ion Chambers



**Gas selection:**  
 < 5 KeV: He  
 5 – 15 KeV: N2  
 > 15 keV: Ar

Current  
(nA to  $\mu$ A)



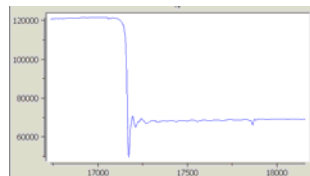
Current  
amp:  
outputs  
voltage



voltage



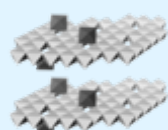
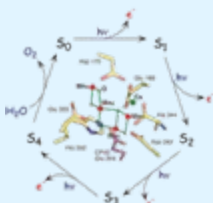
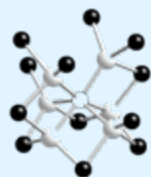
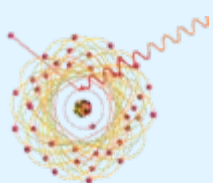
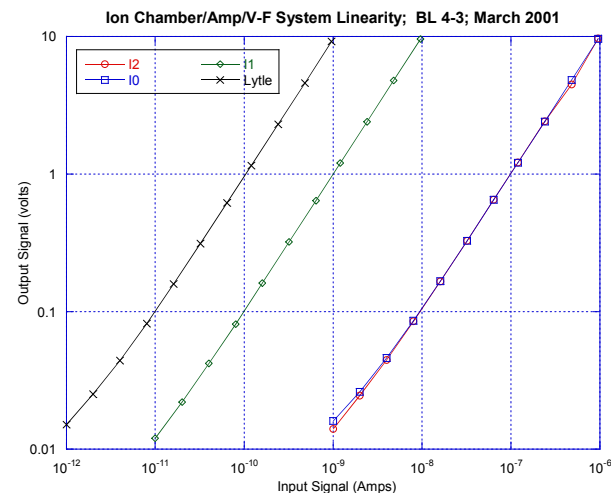
V-F  
converter:  
Pulses to  
computer



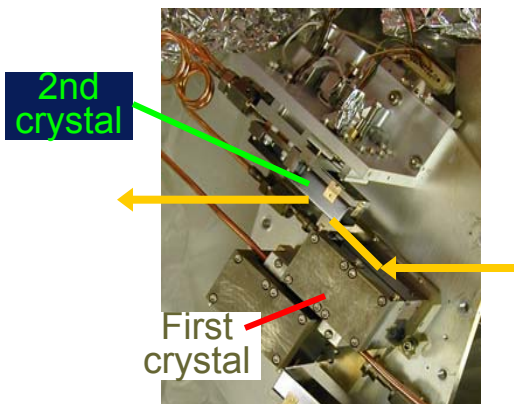
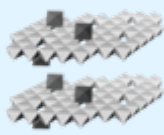
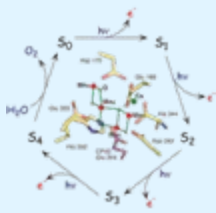
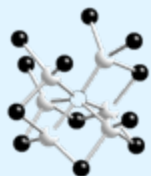
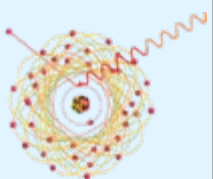
**Linear range:** up to 5 V (SR570) or 10 V (Keithley)

**Linear range:** 0.1 to 10 V

**Maintain signal  
below ~ 5 V!!!**



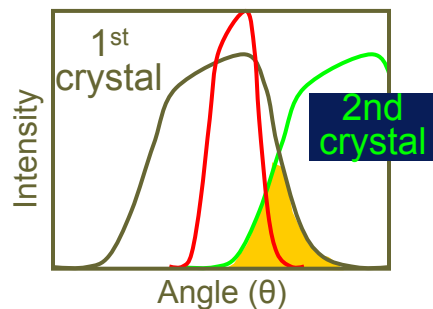
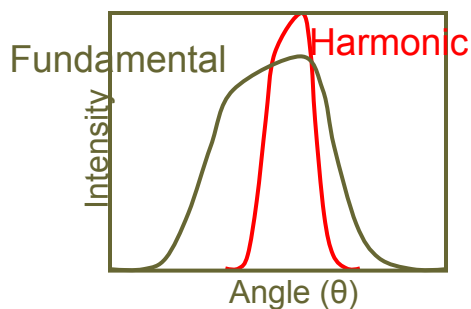
# I. Beam line set-up and optimization: Monochromator tuning



**Bragg's law:**  
 $n \cdot \lambda = 2 \cdot d \cdot \sin(\theta)$

n=1 : "fundamental"

n>1 : "harmonic"



**"Detuning"**: rotating 2<sup>nd</sup> crystal slightly away from diffraction condition.

→ Reduces contribution from harmonics!

→ Typical values:

~40% @ 6 keV

~25% @ 13 keV

~15% @ 20 keV



# I. Beam line set-up and optimization: Choice of monochromator crystal

Si(220): Energy range: ~4 to 40 keV, higher E resolution

Si(111): Energy range: ~2 to 20 keV, lower E resolution

Structural Molecular Biology  
**SMB/XAS**  
SSRL  
X-ray Absorption Spectroscopy

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INSTRUMENTATION  
SOFTWARE  
SUPPORT  
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Stanford Linear Accelerator Center

### SSRL SMB/XAS Interactive Crystal Glitch Database

Enter an element symbol and edge for a default EXAFS scan range and to return cursor values in  $k$  (units in 1/Å).

Chemical Element:        $E_0$  (keV):   
Edge:         
Note: The dashed line denotes the chosen element's edge energy.      Max  $k$  Value to plot:      

Display Options:

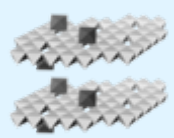
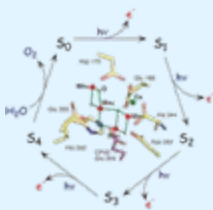
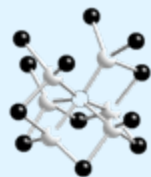
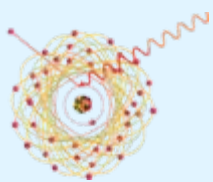
Trace 1:      

Trace 2:      

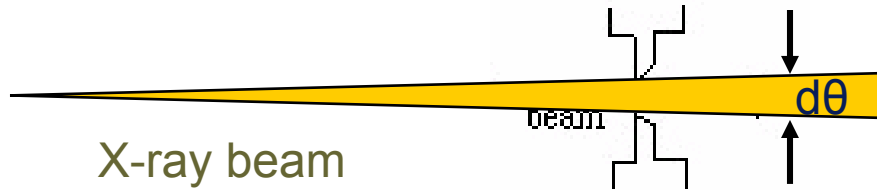
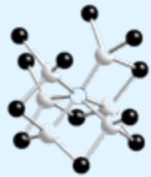
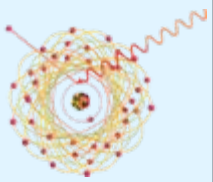
Trace 3:      

Trace 4:      

Choose new energy range:  
 to  keV



# I. Beam line set-up and optimization: Slits control energy resolution!

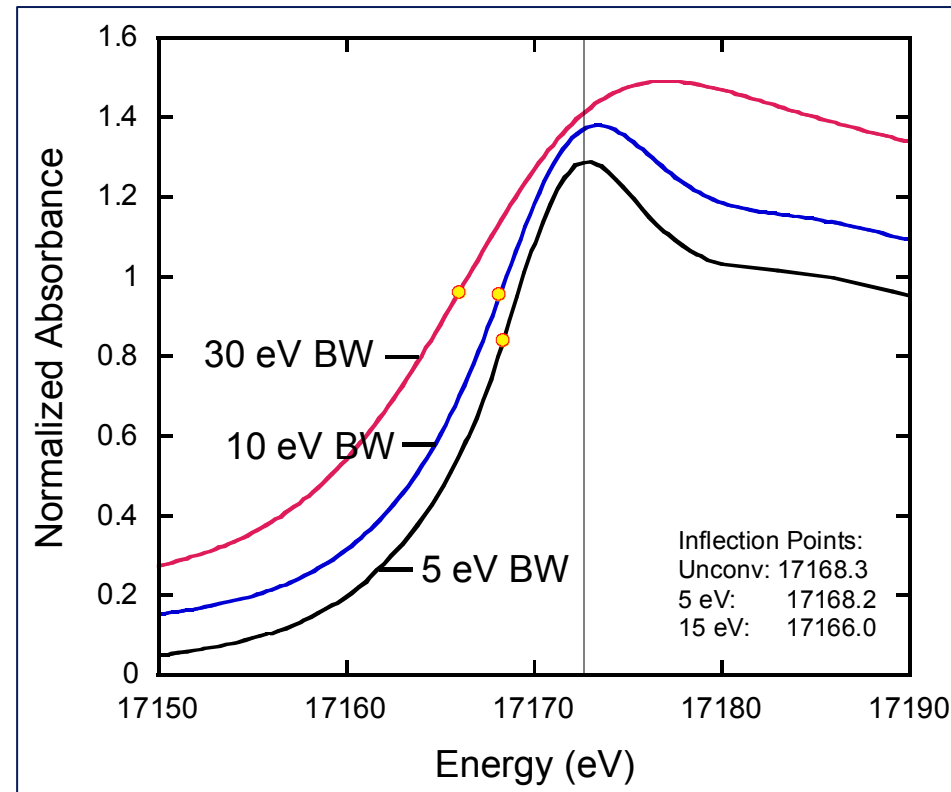
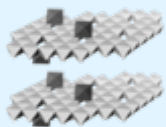
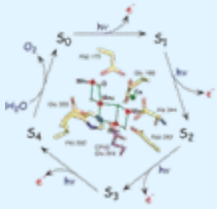


**E resolution,  $dE/E$**   
 $= d\theta / \text{Tan}(\theta_{\text{bragg}})$

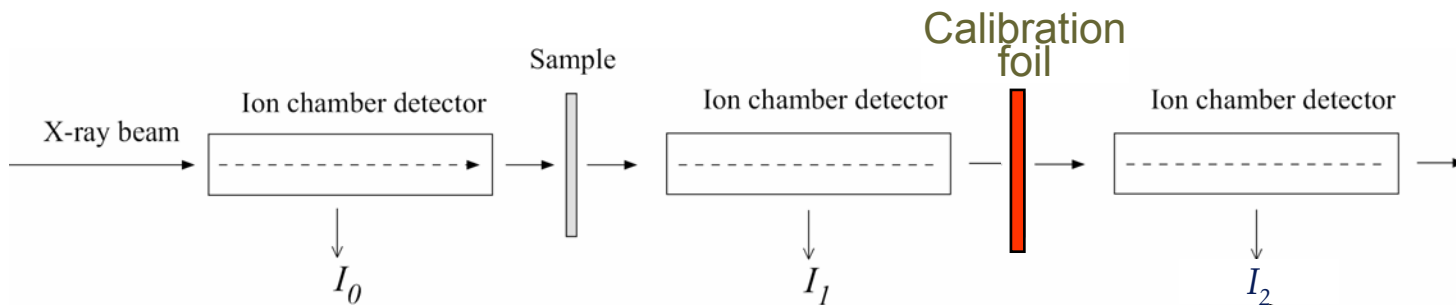
To change energy resolution: Change slit opening.

*Big effect on edge shape AND apparent calibration!*

**ALSO – choice of crystal (e.g. (220) vs (111)) impact energy resolution**



# I. Beam line set-up and optimization: Mono energy calibration

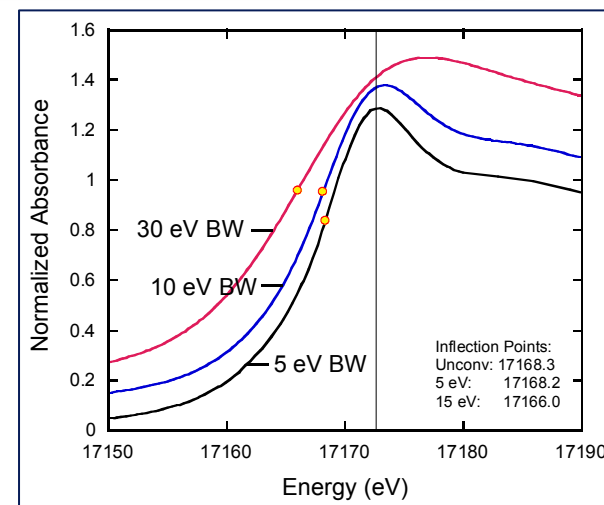


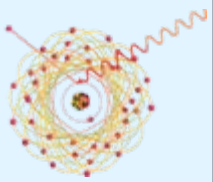
Calibration foil located between  $I_1$  &  $I_2$ .

**Remove foil** after taking calibration (check calibration between every other sample).  
- OR – use calibration foil **different** from sample element (continuous calibration).

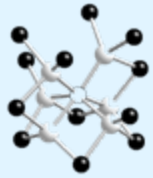
Calibrate on first inflection point of rising edge (preferred) or on top of white line.

**Use consistent energy resolution!**: mono crystal, same slit opening. Good strategy: close slits so spectrometer resolution is < core hole life time.

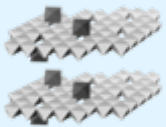
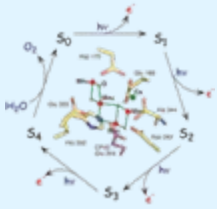


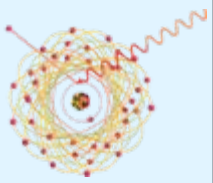


## II. Sample alignment and detectors



- Transmission vs fluorescence geometry
- Transmission geometry
- Lytle detectors for fluorescence yield detection
- Ge detector: highly dilute, chemically complex samples





## II. Sample alignment and detectors: Transmission vs. fluorescence geometry

	Advantages	Requirements	Comments
<b>Transmission mode with ion chambers</b>	Simple Collect 100% of signal No count-rate limitation	Constant sample density, thickness!!!! Concentrated, rel. pure samples.	Eliminate harmonics!
<b>Fluorescence mode with ion chambers</b>	Simple Collect ~10% of sphere No count-rate limitation	> 500 ppm No strong interfering elements	Beware: over-absorption!
<b>Fluorescence mode with energy-dispersive detectors</b>	Excellent for dilute samples or samples with interfering fluorescence lines	< 300 KHz count rate total count rate	Beware: over-absorption and dead-time!

### Examples:

**Concentrated solids:** grind to fine powder, thoroughly mix with BN or  $\text{LiCO}_3$ , use trans mode.

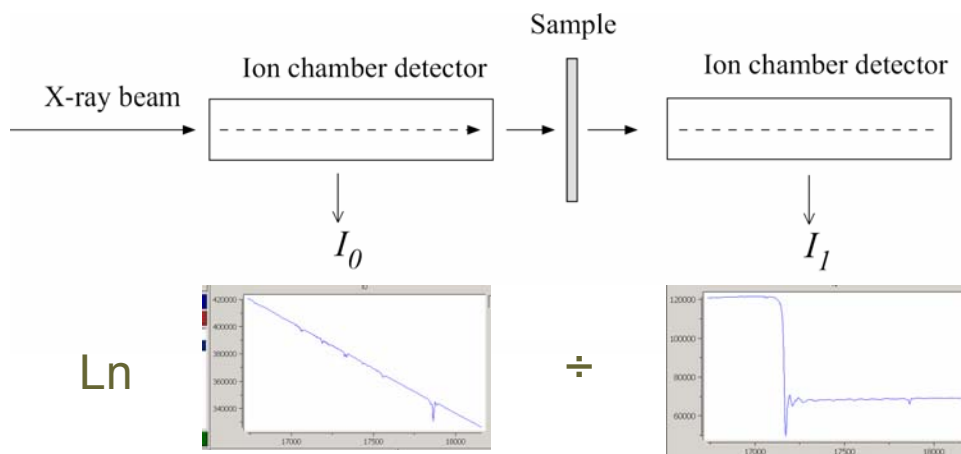
**Thick solid suspensions:** run in trans if mechanically stable and can make thin enough.

**Aqueous solutions:** typically fluo mode with E-disp. detector. Maximize concentration.

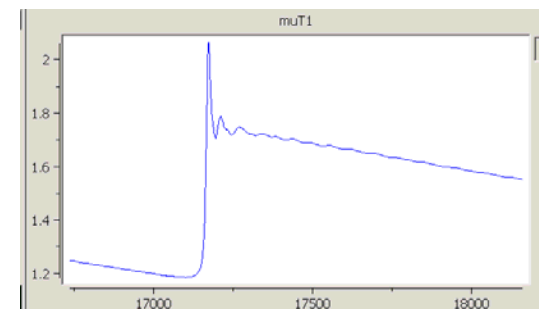
**Soils:** fluo mode typically with E-disp. detector



## II. Sample alignment and detectors: Transmission geometry



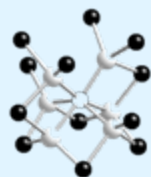
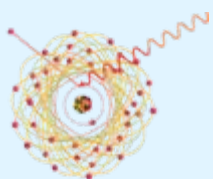
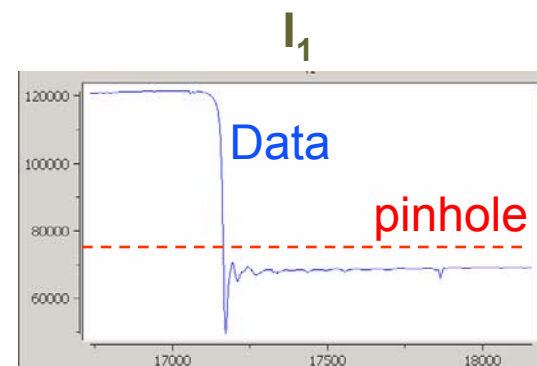
Beer's law:  
Absorbance  $\sim (\ln I_0/I_1)$



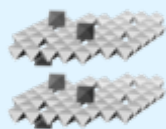
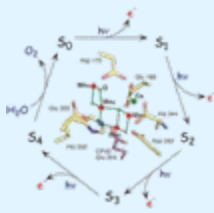
**Beware**,  $I_0$  and  $I_1$  can contain “junk”  
intensity not proportional to EXAFS:  
*e.g.*,

$I_1 = \text{data} + \text{pinhole intensity} + \text{harmonics}$   
 $+ \text{dark current}$

When junk intensity  $\sim$  data then spectra  
will be screwed up!



Ln



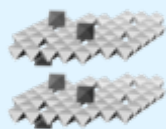
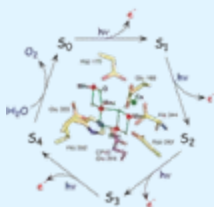
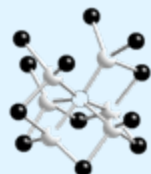
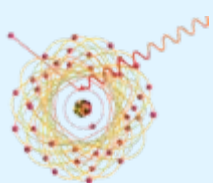
## II. Sample alignment and detectors: Transmission geometry

*And, your data will look like:*



**Rules for transmission samples:**

- Must be homogeneous on  $1 \mu\text{m}$  scale
- Use small slits –typically NOT count-rate limited!
- Must be of rigorously constant thickness
- Must rigorously eliminate harmonics
- Must measure/subtract dark current
- Ideal sample:  $I_1$  drops by 70 to 90% over edge

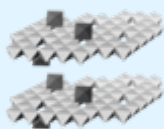
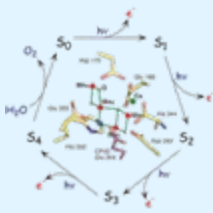
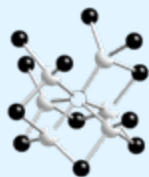
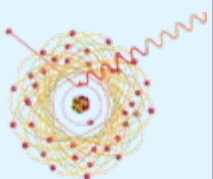


## II. Sample alignment and detectors: How to prepare transmission samples

Ideally, wish to prepare powder samples that have the same homogeneity of a  $\sim 2 \mu\text{m}$ -thick metal foil!

How do we do this in a sample that is typically  $\sim 1 \text{ mm}$  thick?

- *Proper density* – achieved by mixing small quantity of sample into a weakly-absorbing matrix.
  - Typical matrices: BN, sucrose,  $\text{Al}_2\text{O}_3$ .  *$\text{Al}_2\text{O}_3$  is often best because it is not redox active and it is very hard, so it can be used to further mill the sample.*
- How much compound to add? – Can be calculated using web tools at <http://www.cxro.lbl.gov/> to obtain  $\sim 80\%$  absorption by the metal of interest above the edge. Typical ratio is 20 mg of sample in 70 mg of BN or  $\text{Al}_2\text{O}_3$ .
- *Homogeneity* – is achieved by first milling your sample and matrix separately and thoroughly using mortar/pestle to obtain particle size  $< 1 \mu\text{m}$ . Then, weigh sample into matrix and continue to mix
- Must be of rigorously constant thickness: load into stiff sample holders.
- Pressing pellets is helpful, but beware of preferred particle orientation!



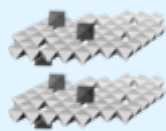
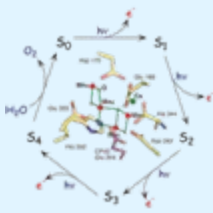
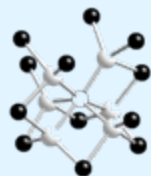
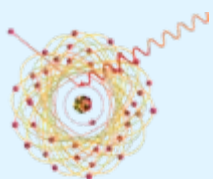
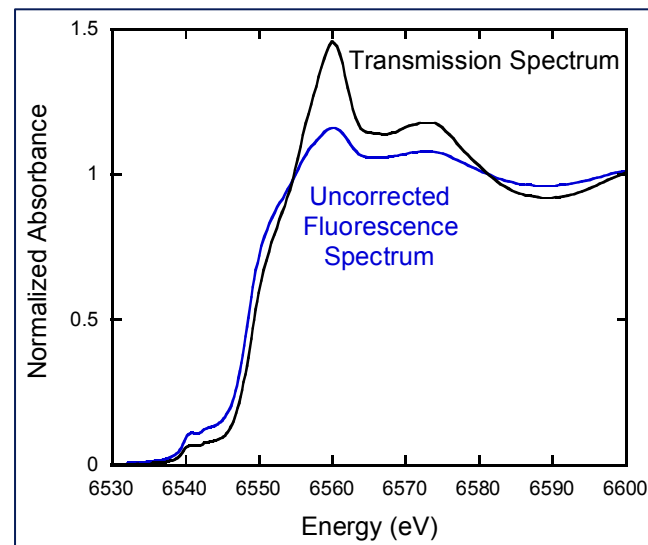
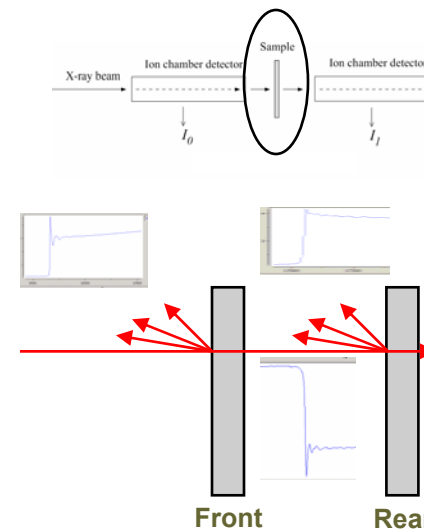
## II. Sample alignment and detectors: Fluorescence geometry

**Dilute sample paradigm** – assumes absorption of beam is so weak that it does not corrupt amplitudes from rear of sample.

Concentrated samples will suffer amplitude reduction, so called, “**over-absorbance**” effect.

Can strongly modify XANES region.

Mitigation: run concentrated samples in transmission, with electron yield. In some cases, it is possible to analytically correct for self absorbance (Corwin Booth’s talk this AM.



## II. Sample alignment and detectors: Lytle detector

Good for relatively pure and moderately dilute samples (~1,000 to 20,000 ppm range).

Ionization chamber detector: *no* practical count rate limit

Gases: Ar (< 10 KeV), Xe (10 – 15 keV), Kr (>15 keV) – energies of *emission* lines!

Use **x-ray filters** in conjunction with Soller slits to reduce elastic scattering from signal.

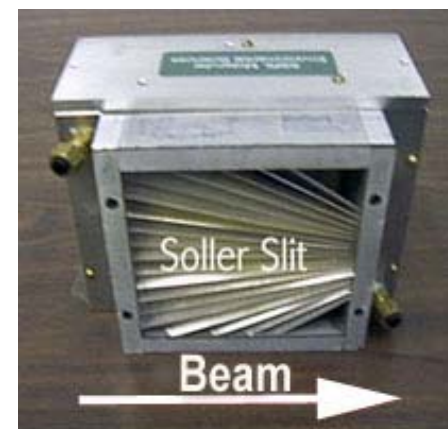


Figure 2. Soller Slit Position

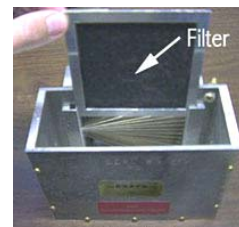
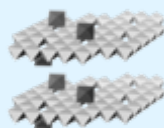
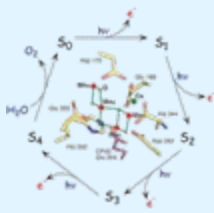
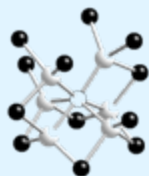
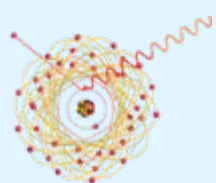
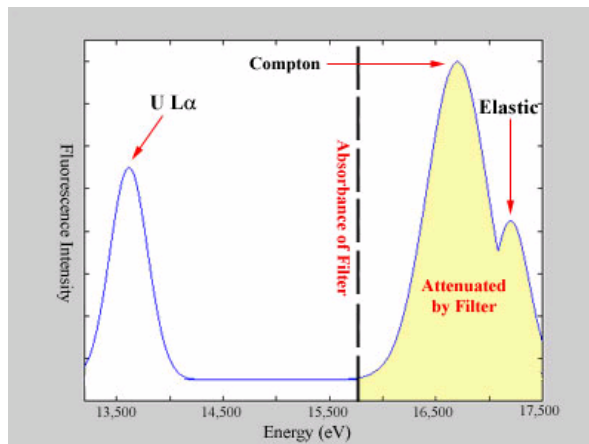
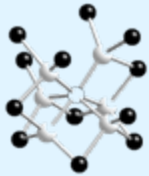
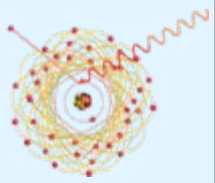


Figure 3. X-ray Filter Position

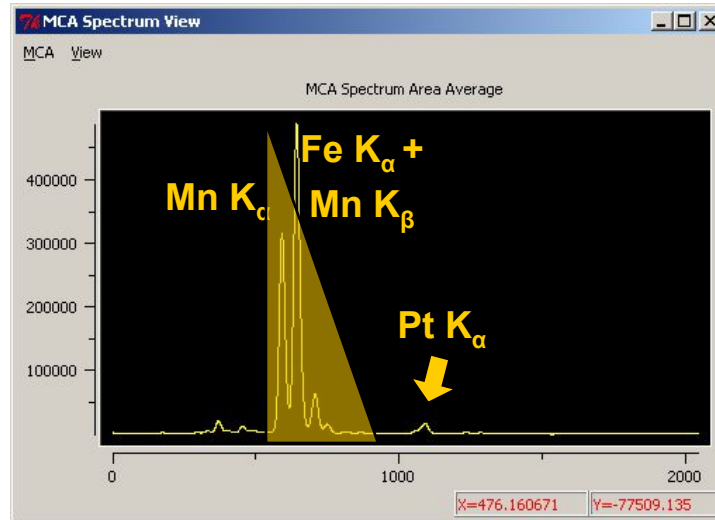
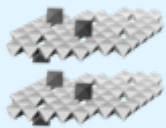
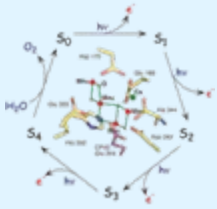




## II. Sample alignment and detectors: Dilute & chemically heterogeneous samples



Pt in marine  
ferro-  
manganese  
crusts

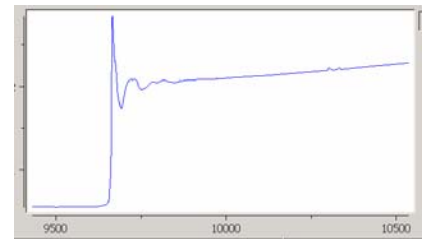
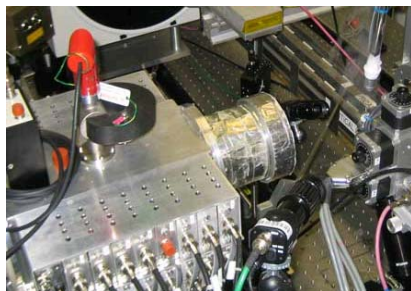
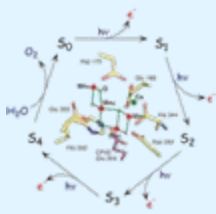
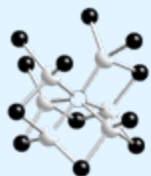
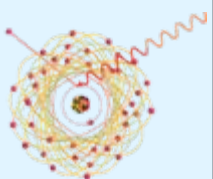


**Solid-state detectors** (single Ge and Si crystals) provide energy resolution of *ca* 250 eV FWHM and can resolve individual emission peaks.

**Disadvantage:** count-rate limitation to  $\sim 280,000$  counts/sec

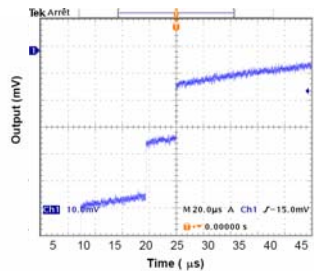
Use high-pass **x-ray filters** (in this case, V or Al) to cut “background” counts and thus allow for more Pt counts.

# II. Sample alignment and detectors: Solid state detectors: basics



**Preamp**

Voltage ramp



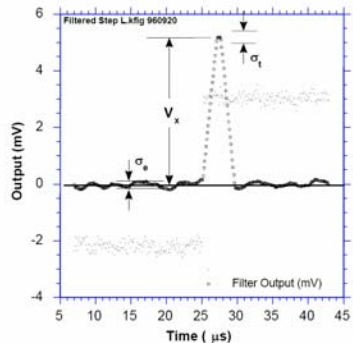
**Incoming Count Rate, "ICR"**  
~1,000,000 cps

**Windowed or "SCA" counts,**  
< 600,000 cps

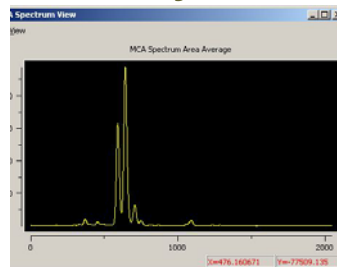


Voltage pulse train

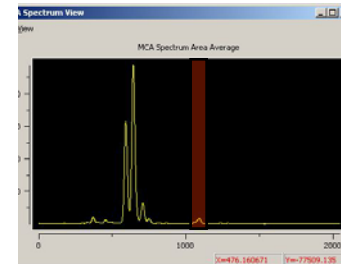
**Amplifier**



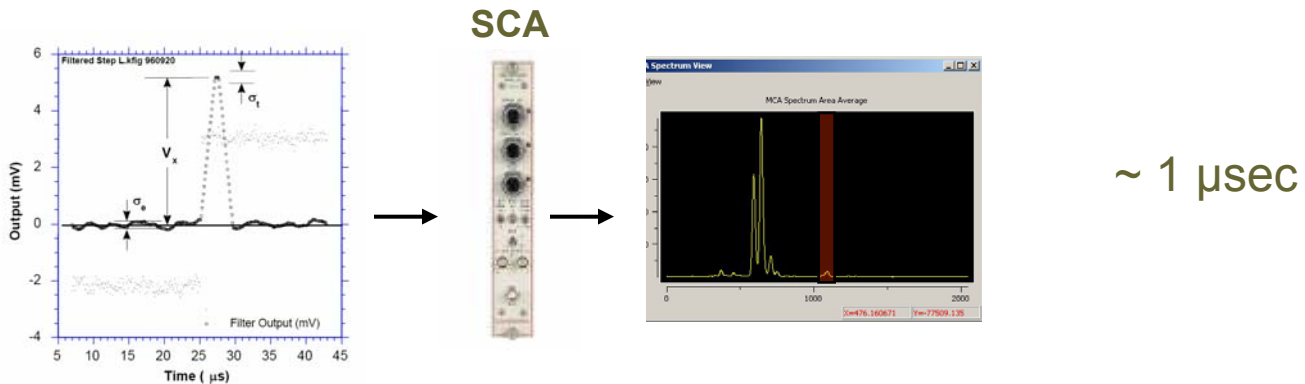
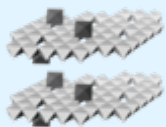
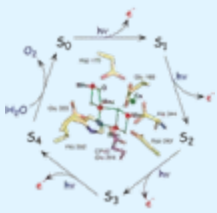
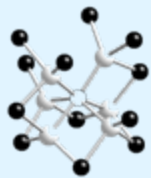
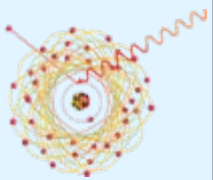
**Multichannel analyzer**



**SCA**



## II. Sample alignment and detectors: Solid state detectors: basics

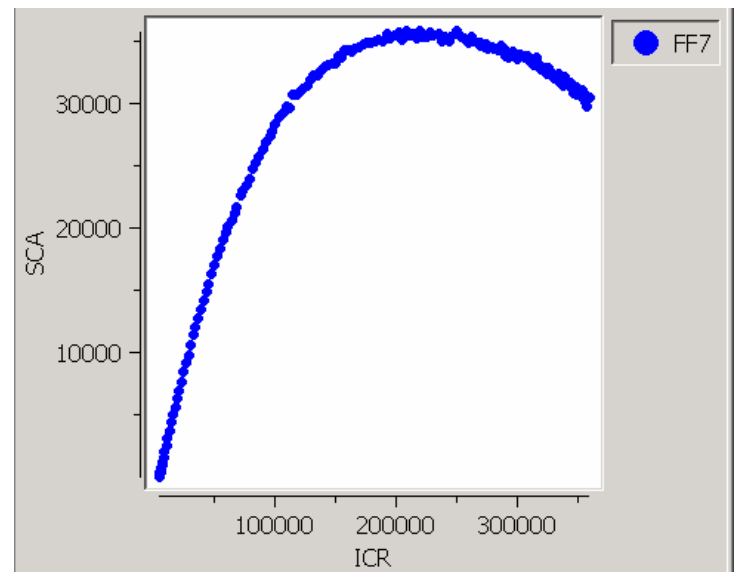


When count rate approaches  
~100,000 counts/sec, detector  
becomes paralyzed during  
some events = "deadtime",  
according to:

$$SCA = K \cdot ICR_t \cdot \exp(-ICR \cdot T_d)$$

$K$  = constant,  $T_d$  = dead time.

Data can be quantitatively  
corrected (hands on sessions)



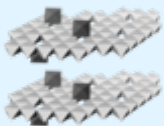
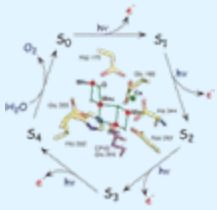
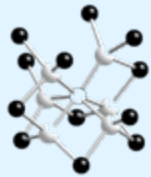
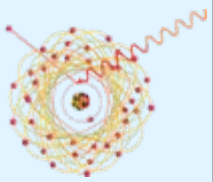
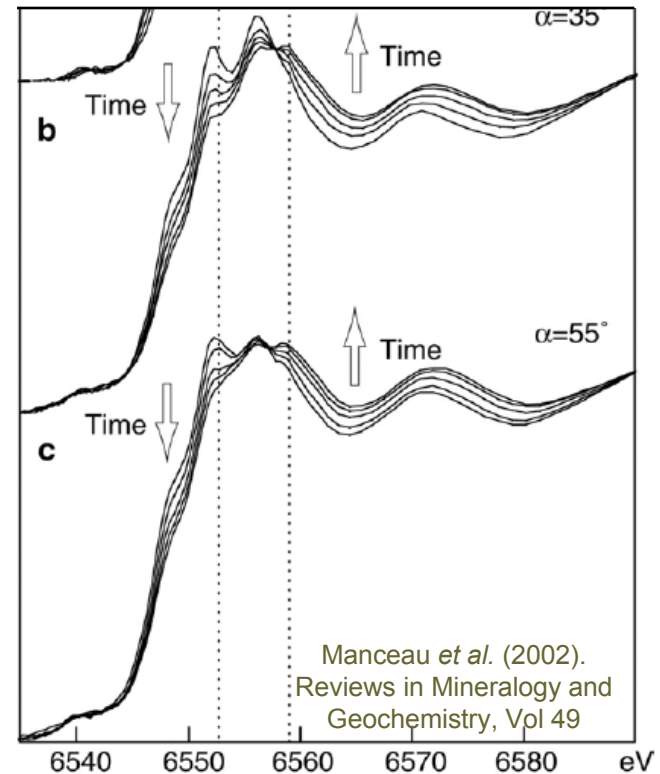
# III. Data acquisition

## To be discussed during hands-on sessions:

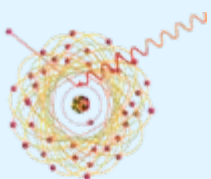
- Setting up regions files - optimizing counting time, data range
- How to check data quality
- What will be the good data range?
- How many scans are enough?

## Beam damage...

some samples are particularly subject to photo-induced redox changes. Mitigation: typically cryogenic temperature for data acquisition.

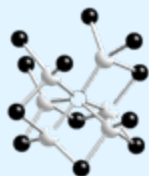


# III. Data acquisition which beam line should I use?



**Model compounds** (concentrated, compositionally simple):

BL: 4-1, 4-3, 10-2



**Moderately dilute samples:**

BL: 4-1, 4-3, 10-2

**Highly dilute and/or chemically heterogeneous samples:**

BL: 7-3, 9-3, 11-2

**Low-energy XAS (~2.1 - ~6 keV):**

BL: 6-2, 4-3

**High-energy XAS (~17 - 38 keV):**

BL: 4-1, 7-3, 10-2, 11-2

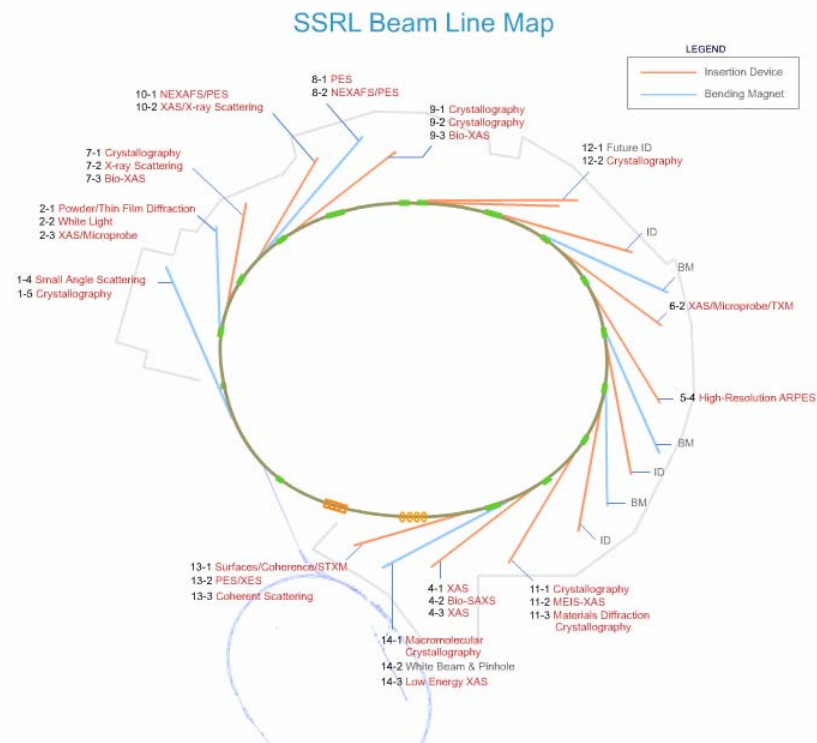
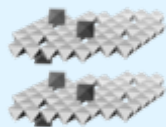
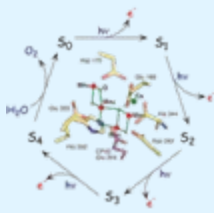
**Mat Sci:** BL: 4-1, 4-3, 10-2

**Environmental:** BL: 4-1, 4-3, 10-2, 11-2

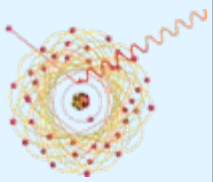
**Biological:** BL: 7-3, 9-3

**Micro-XAS:** BL 2-3

**Grazing-incidence XAS:** BL 11-2

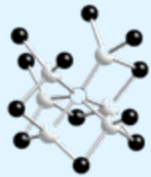






# Quiz!

Connect the pictures to the data...



**Bragg's law:**  
 $n \cdot \lambda = 2 \cdot d \cdot \sin(\theta)$

